Musashi Binding Elements in Zika and Related Favivirus 3'UTRs

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Background and outline

Emerging and re-emerging arthropod-borne viruses such as Japanese encephalitis virus (JEV), Dengue virus (DENV), Yellow fever virus (YFV), and Chikungunya virus (CHIKV) are a growing global health threat. **Zika virus** (ZIKV) is a neurotropic flavivirus (FV) that can cause **con**genital infection, which can result in microcephaly and fetal demise. Recently, the translational regulator protein Musashi-1 (Msi1) has been attributed to promoting ZIKV replication, neurotro**pism, and pathology** [1]. Msi1 predominantly binds single-stranded UAG motifs in the 3'UTR of RNA [2]. Here we systematically analyzed the thermodynamic properties of Musashi binding elements (MBEs) in the 3'UTR of 76 arbovirus genomes in silico. Our results indicate that MBEs in the ZIKV 3'UTR occur predominantly in unpaired, single-stranded structural context, thus corroborating experimental observations of Msi1 binding affinity with a thermodynamic model of RNA structure formation.

Opening energy and single-strandedness

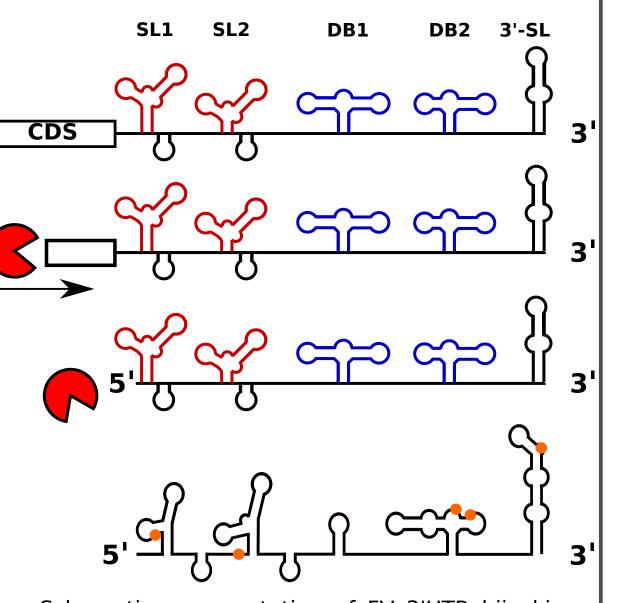
We use the ViennaRNA Package [3] to model the thermodynamics of RNA secondary structure formation. The partition function Z allows for computation of the equilibrium probability of secondary structure s

$$Z = \sum_{s} e^{-E(s)/RT} \qquad P(s) = \frac{e^{-E(s)/RT}}{Z}$$

The **accessibility** (i.e. the probability that a region $i \dots j$ along the RNA

Flavivirus 3'UTR mediates pathogenicity

Flaviviruses are small (+)ssRNA viruses of 10-12kB length with 220highly structured UTRs. Upon in- 5' fection, accumulation of stable long non-coding viral RNA, subgenomic flaviviral RNA (**sfRNA**), is observed. sfRNAs can modulate cellular function and are linked to pathogenicity. They are stable decay intermediates produced by partial degradation of the viral genome by 5'-3' exoribonuclease

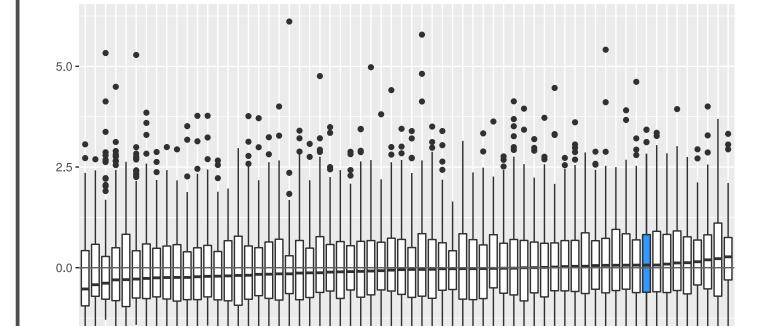


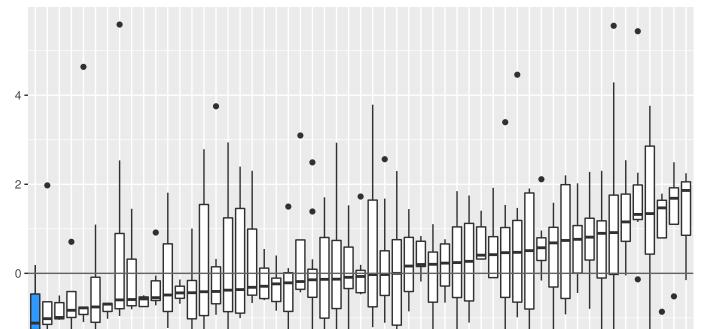
is single-stranded) can be derived from Z. The **opening energy** (i.e. the free energy required to force the region to be in a single stranded structural context) is computed as $\Delta G_{\text{open}} = -RT \ln P(\text{unpaired})$. Low opening energy indicates single-strandednesss. We compute local pairing proba- $z = \frac{\Delta G_{\text{open}}(x) - \mu}{z}$ bilities of trinucleotides to assess the likeli-

hood of MBE single-strandedness in a genomic context. Comparison to a large sample of randomized sequences allows to compute a zscore for each trinucleotide.

MBEs are highly accessible in ZIKV 3'UTR

We analyzed the accessibility of all trinucleotides in the coding region (CDS) and 3'UTR of ZIKV from Brazil and found a marked difference in the distribution of z scores, suggesting different sequence composition. Musashi-binding **uag trinucleotides are maximally accesible** in the 3'UTR, which corroborates experimental studies [1,4].





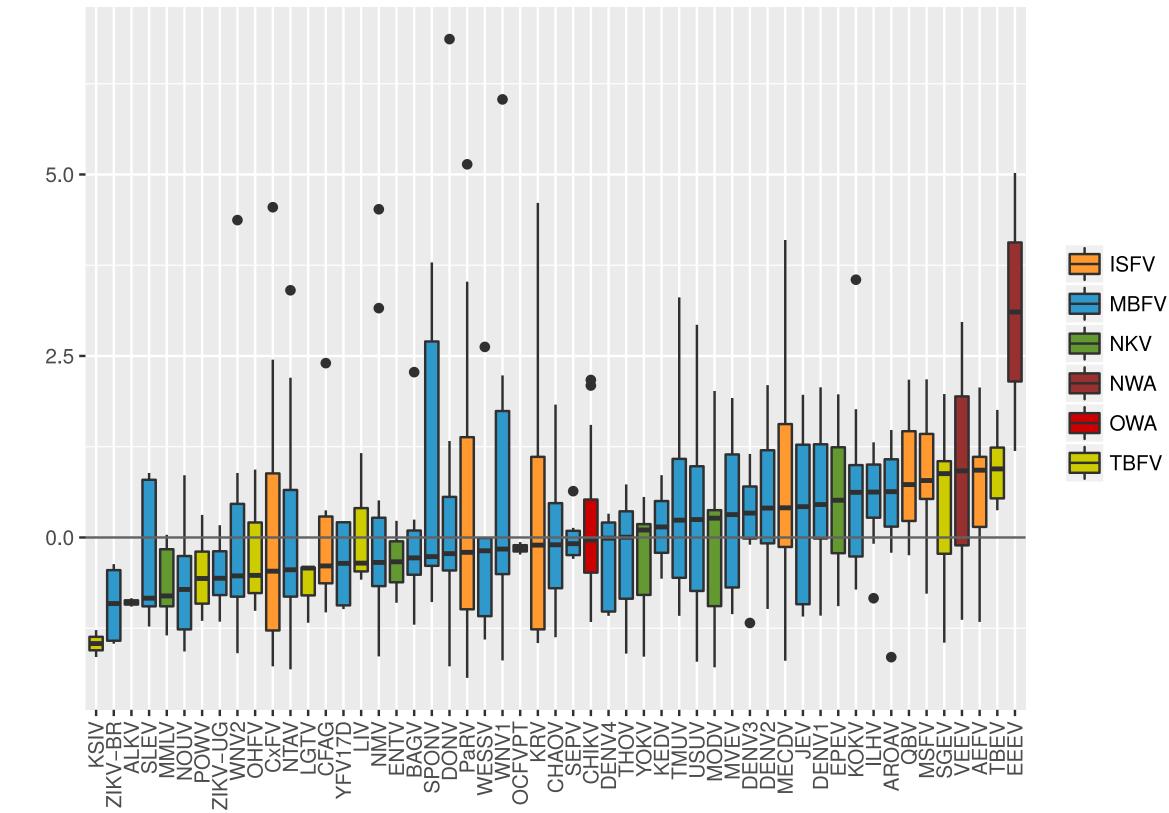
Xrn1, which is efficiently stalled at the host mRNA degradation pathway. Conserved xrRNA stemevolutionary conserved xrRNA tandem copies within 3'UTRs and stall the host exonuclease structures.

Fig 1. Top: Schematic representation of FV 3'UTR hijacking loop (SL) and dumbbell (DB) elements are located in single or Xrn1 (red pac-man). Bottom: ZIKV has two SL and one DB element. Location of UAG MBEs are highlighted in orange.

Fig 2. Distribution of z scores for all trinucleotides in the CDS (left) and 3'UTR (right) of ZIKV-BR, sorted by median z score. Interquartile ranges are homogeneous within the CDS region. The MBE motif UAG (blue) is maximally accessible in the 3'UTR.

MBE accessibility in related viruses

To address the broader question whether other viruses have a similar neurotropic potential to ZIKV in the developing fetus, we analyzed MBEs in the 3'UTR of related arbovirus genomes. Low overall z scores indicate unpaired UAG motifs, suggesting high Msi1 affinity. We therefore use it as an **estimator for teratogenicity**.



Conserved xrRNAs contain MBEs

We localized MBEs in the 3'UTR of FV and found a perfectly conserved use trinucleotide pair in the dumbbell (DB) element. Both MBEs appear in an unpaired structural context, rendering them perfect targets for the two Musashi RNA recognition motif domains.

	(((((((((((((()))))))))))	((()))))))))))	
WNV.01	GCGCAGC	CAUAAC	CUUGGUGA-	AGGUGUUA	GCCAA <mark>G</mark>	G AGAAG <mark>G</mark>	GACUAGA	GGUU	AGCAGA <mark>G</mark>	AUCCU	GCGC	69
KUNV.02	ACGCGGC	CCUAGC	UCUGGCAA-	UGGUGUUA2	ACCAG <mark>A</mark> (J UGAAA <mark>G</mark>	GACUAGA	GGUU	AGAGGA <mark>G</mark>	ACCCC	GCGU	69
APCV.03	CCGCGGC	CCAACC	AGUUCAGA	CU-GAUGCUAI	UGAAC <mark>U</mark> (GGUAA <mark>G</mark>	GACUAGA	GGUU	AGAGGA <mark>G</mark>	ACCCC	GCGG	70
JEV.04	CCACGGC	CCAAG <mark>C</mark>	UUCGUCUA-	-G-GAUGCAAUA	GACGA <mark>G</mark>	G UGUAA <mark>G</mark>	GACUAGA	GGUU	AGAGGA <mark>G</mark>	ACCCC	GUGG	71
JEV.05	CCACGGC	CCAAG <mark>U</mark>	CUCGUCCA-	-G-GAUGCAAUG	GACGA <mark>G</mark> Z	A UGUAA <mark>G</mark>	GACUAGA	GGUU	AGAGGA <mark>G</mark>	ACCCC	GUGG	71
JEV.06	CCACGGC	CCAAAC	CUCAUCUA-	-G-GAUGCAAUA	GAUGA <mark>G</mark>	G CGUAA <mark>G</mark>	GACUAGA	GGUU	AGAGGA <mark>G</mark>	ACCCC	GUGG	71
WNV.07	ACGCGGC	CCAAA <mark>U</mark>	CCUGGUGA-	-U-GGUGUUA-U	GCCAG <mark>G</mark>	G UGGAA <mark>G</mark>	GACUAGA	GGUU	AGAGGA <mark>G</mark>	ACCCC	GCGU	70
MVEV.08	CCGCAGC	CCGGG <mark>C</mark>	CGGGAGGA	GGUGAUGCGA-A	C-CCC <mark>G</mark>	GC-GAA <mark>G</mark>	GACUAGA	GGUU	AGAGGA <mark>G</mark>	ACCCU	GCGG	70
ALFV.09	CCACGGC	CCGGGC	CAUGAGU-(GAUGAUGUUA-A	CUCAUG	GC-GAA <mark>G</mark>	GACUAGA	GGUU	AGAGGA <mark>G</mark>	ACCCC	GUGG	70
USUV.10	CCACGGC	UCAAG <mark>C</mark>	GAACAGAC(GGUGAUGCGA-A	CUGUUC	J UGGAA <mark>G</mark>	GACUAGA	GGUU	AGAGGA <mark>G</mark>	ACCCC	GUGG	72
USUV.12	UCAC GGC	CCAAGC	GAACAGAC(GGUGAUGCGA-A	CUGUUC	J UGGAA <mark>G</mark>	GACUAGA	GGUU	AGAGGA <mark>G</mark>	ACCCC	GUGG	72
SLEV.13	CG GCGGC	CCAAAC	CAUGGAG		-CCAU <mark>G</mark>	G CGUAA <mark>G</mark>	GACUAGA	GGUU	AGAGGA <mark>G</mark>	ACCCC	GCUG	66
SLEV.14	CAGC GGC	CCAAAC	CAUGGAG	UGCGUGA	-CCAU <mark>G</mark>	G CGUAA <mark>G</mark>	GACUAGA	GGUU	agagga <mark>g</mark>	ACCCC	GCUG	66
											70	
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Fig 4. Structural alignment of DB elements in the 3'UTR of Japanese encephalitis group viruses. Nucleotide coloring indicates covariation. Conserved UAG trinucleotides are found in the cental multiloop and the distal hairpin loop (blue boxes). Bars below the alignment indicate+++ the level of sequence conservation.

Fig 3. MBE opening energy z scores in the 3'UTR of flaviviruses and alphaviruses. Color indicates virus group (ISFV: Insect-specific flaviviruses; MBFV: Mosquito-borne flaviviruses; NKV: No known vector flaviviruses; NWA: New World alphaviruses; OWA: Old World Alphaviruses; TBFV: Tick-borne flaviviruses).

The Brazilian ZIKV isolate has the lowest median MBE opening energy z scores among mosquito-borne FV, followed by the neurotropic viruses SLEV, WNV, and the tick-borne POWV, which can cause transplacental infection, severe neuropathology and fetal demise [5].

Conclusion

We employed an established biophysical model of RNA structure formation to analyze thermodynamic properties of MBEs in silico. Our results underline experimental studies suggesting that ZIKV is not alone in its capacity to cause severe neuropathology [5]. While several tick- or mosquito-borne viral species like Karshi virus (KSIV), Alkhumra hemorrhagic fever virus (ALKV) or Nounané virus (NOUV) line up with ZIKV in our thereretical model, their tropism might have been overseen due to the lack of reported significant outbreaks. However, they appear to have a similar neurotropic potential.

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[4] Zearfoss, N.R. et al. (2014), A conserved three-nucleotide core motif defines Musashi RNA binding specificity, J Biol Chem 289(51):35530-35541 [5] Platt, D.J. et al. (2018), Zika virus-related neurotropic flaviviruses infect human placental explants and cause fetal demise in mice, Sci Transl Med, 10(426) eaao7090.

